



Research Center for Applied Sciences  
Academia Sinica, Taipei, Taiwan

# 體內體外大不同 – 微流體細胞培養

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# Outline

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- **Introduction**
  - **Why Microfluidic Cell Culture?**
- **Microfluidic Cell Culture Devices**
- **Integration**
- **Conclusion**

# 細胞培養... WHY ? ? Problem ? ?

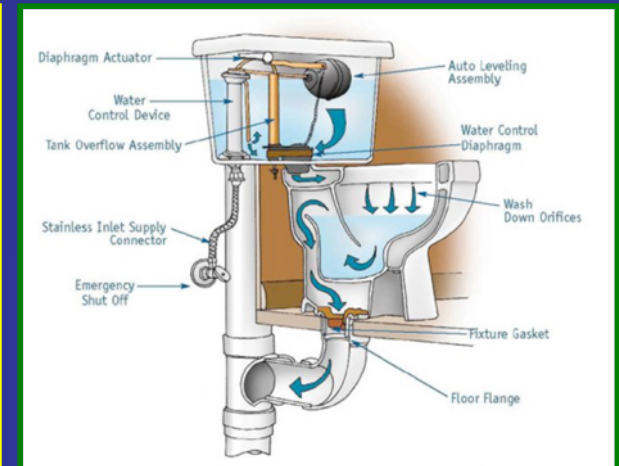
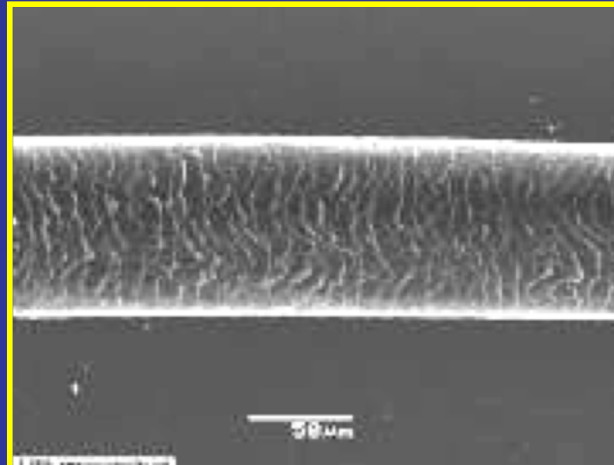
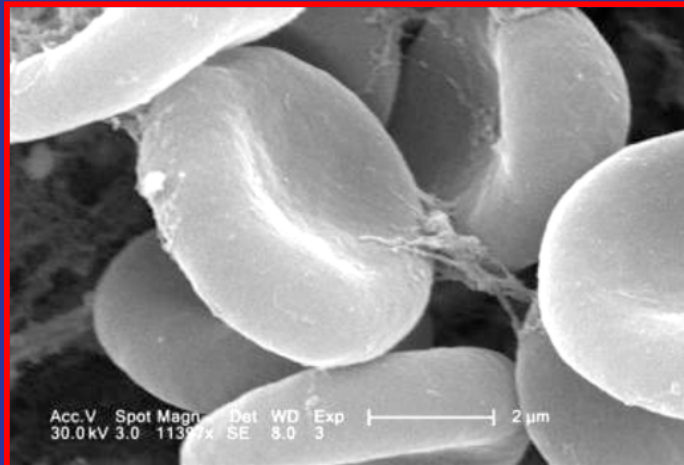
## 細胞培養 (Cell Culture)

*in vivo vs. in vitro*



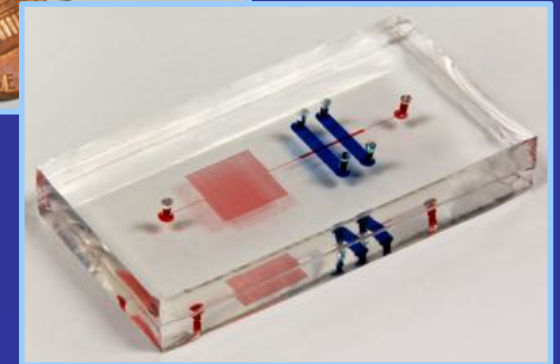
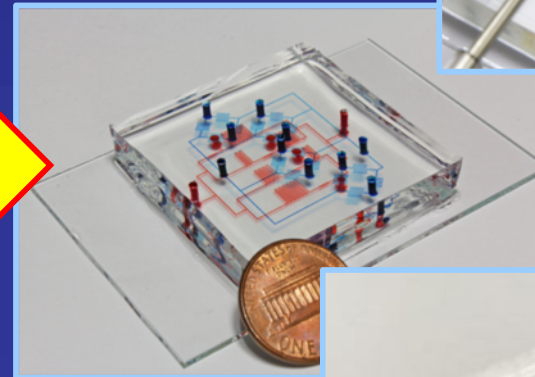
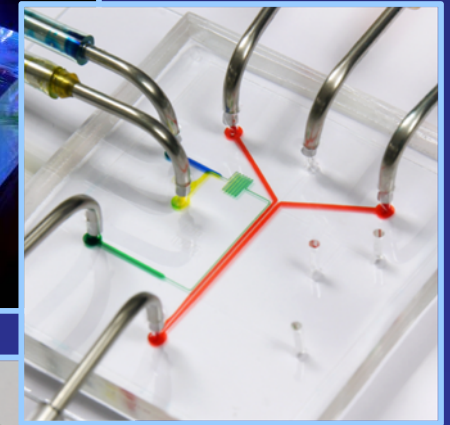
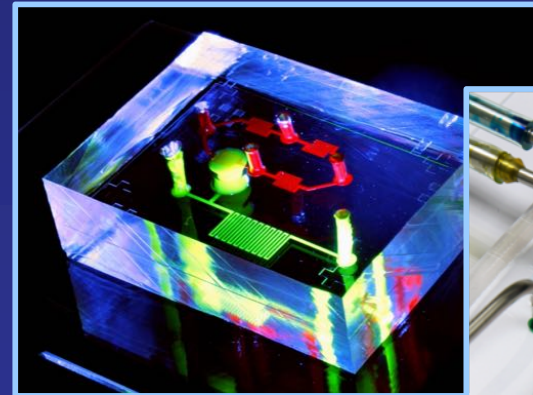
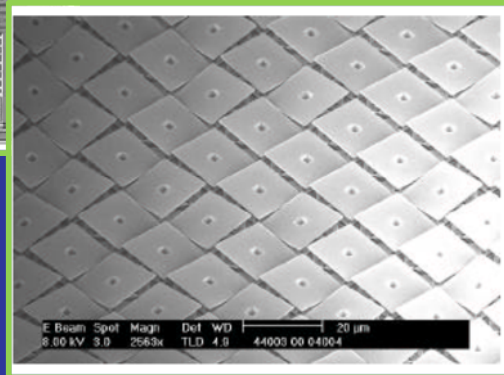
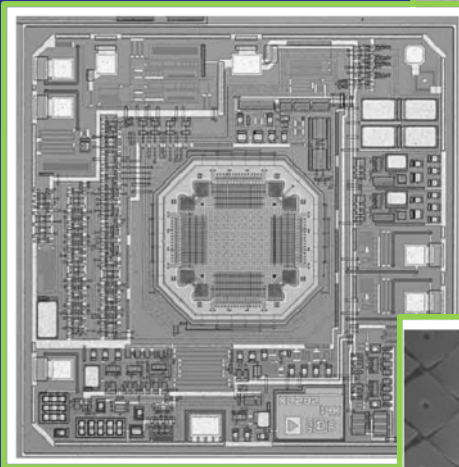
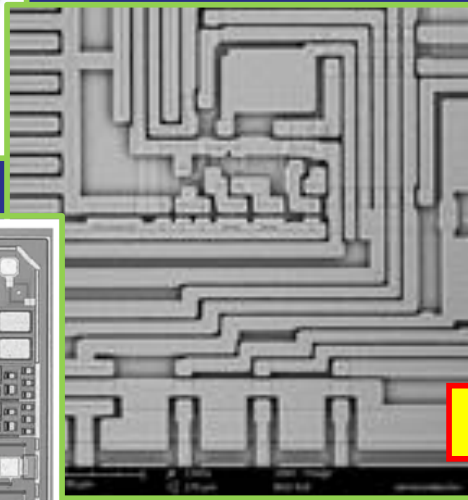
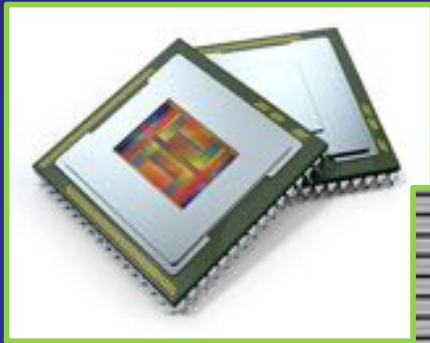
# 微流體 (Microfluidics)··· WHAT ? ?

- Microfluidics = Micro + Fluidics
- Micro/Nano:  $10^{-6}$  ,  $10^{-9}$ 
  - Small size (sub-mm)
  - Small volumes ( $\mu\text{l}$ , nL, pL)
- Fluidics: handling of liquids and/or gases



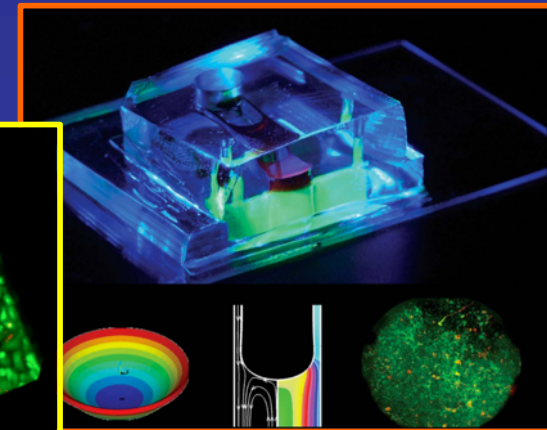
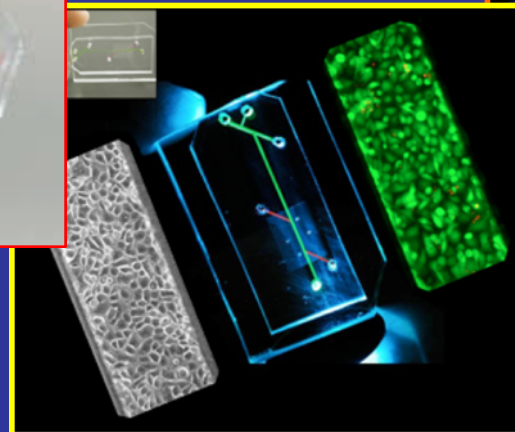
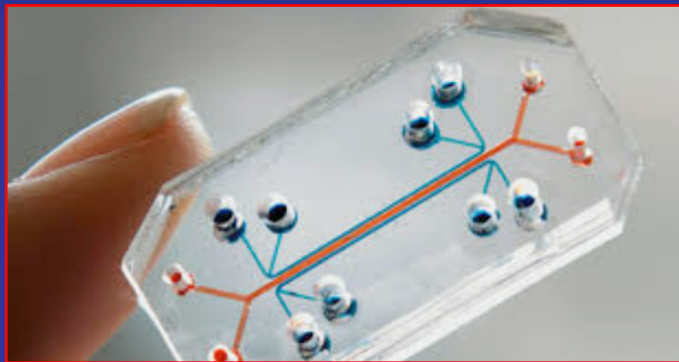
# 晶片 (Chips) ... WHAT??

- 積體電路 (IC), 微機電 (MEMS) 及 微流體晶片



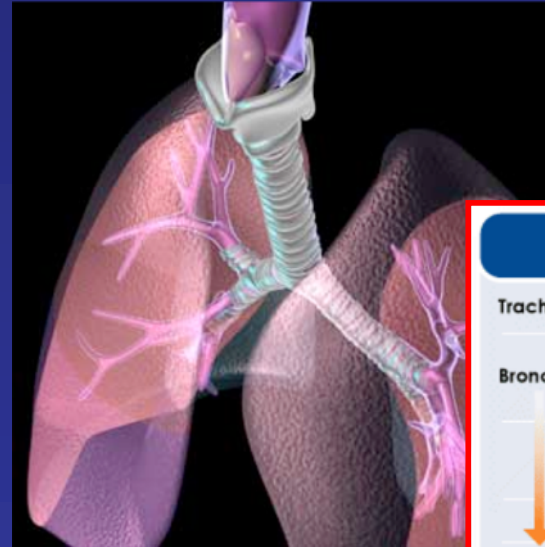
# Introduction – Microfluidic Devices

- **Why Microfluidic Devices?**
  - Unique properties (laminar flow, surface tension...)
  - Small sample volume and easy to scale up
  - Well-controlled microenvironments
  - Precise spatial and temporal control
  - Able to mimic the rich biochemical and biophysical complexity of the cellular microenvironment



# 身體內的微流體系統 *Microfluidics in vivo*

- 循環系統 及 呼吸系統

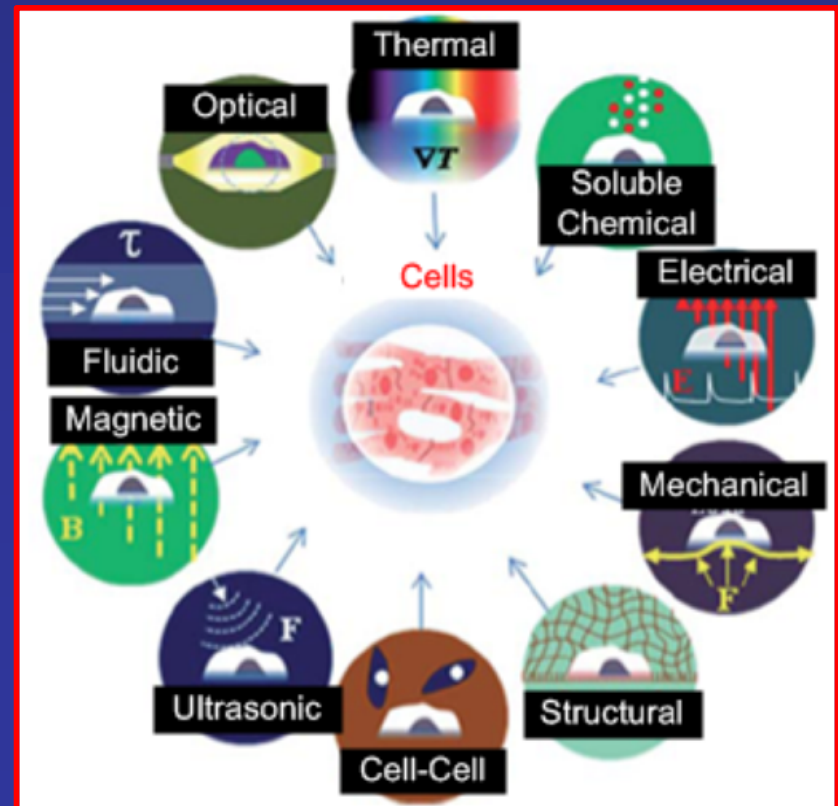


Location	Generation
Trachea	0
Bronchi	1
	2
	3
Bronchioles	4
	5
Terminal bronchioles	↓ 16
	17
Respiratory bronchioles	18
	19
Alveolar ducts	T <sub>1</sub> 20
	T <sub>2</sub> 21
	T <sub>1</sub> 22
Alveolar sacs	T 23

©2008 Teva Specialty Pharmaceuticals LLC<sup>®</sup>

# Introduction – Gaseous Microenvironments

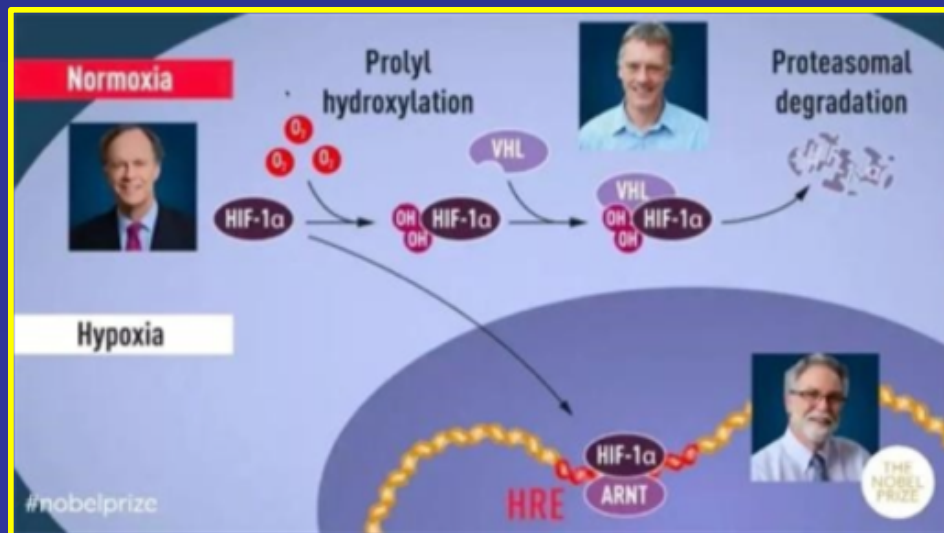
- Why Gaseous Microenvironments?
  - Regulate Important Biological Functions: Breathing, Blood Vessel Dilation etc.
  - Essential Biological Gases: Oxygen, Carbon Dioxide, Nitric Oxide (EDRF) etc.
  - *Important* but *Difficult* to Control.
- Diffusivity in water at 20°C:
  - O<sub>2</sub>:  $1.97 \times 10^{-5}$  cm<sup>2</sup>/s
  - CO<sub>2</sub>:  $1.76 \times 10^{-5}$  cm<sup>2</sup>/s
  - NO:  $2.21 \times 10^{-5}$  cm<sup>2</sup>/s
  - Ribonuclease (14 kDa):  
 $0.12 \times 10^{-5}$  cm<sup>2</sup>/s
  - Albumin (65 kDa):  
 $0.060 \times 10^{-5}$  cm<sup>2</sup>/s





# Introduction – Oxygen Tension

- Oxygen tension plays an essential role in biological systems. For examples:
  - Tumor Malignant Progression and Angiogenesis
  - Cancer Treatment
  - Stem Cell Differentiation



## The Nobel Prize in Physiology or Medicine 2019



III, Niklas Elmedhed. © Nobel Media.  
William G. Kaelin Jr  
Prize share: 1/3



III, Niklas Elmedhed. © Nobel Media.  
Sir Peter J. Ratcliffe  
Prize share: 1/3



III, Niklas Elmedhed. © Nobel Media.  
Gregg L. Semenza  
Prize share: 1/3

The Nobel Prize in Physiology or Medicine 2019 was awarded jointly to William G. Kaelin Jr, Sir Peter J. Ratcliffe and Gregg L. Semenza "for their discoveries of how cells sense and adapt to oxygen availability."

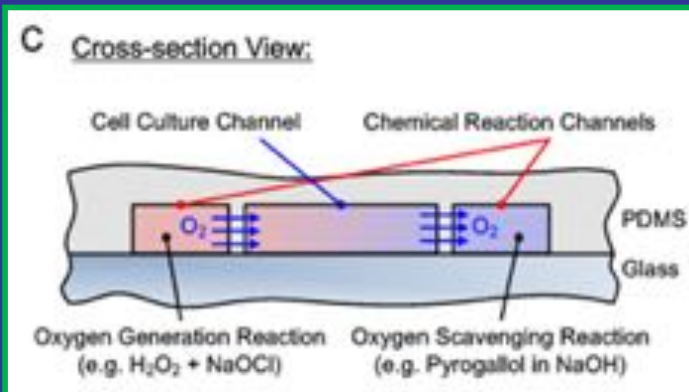
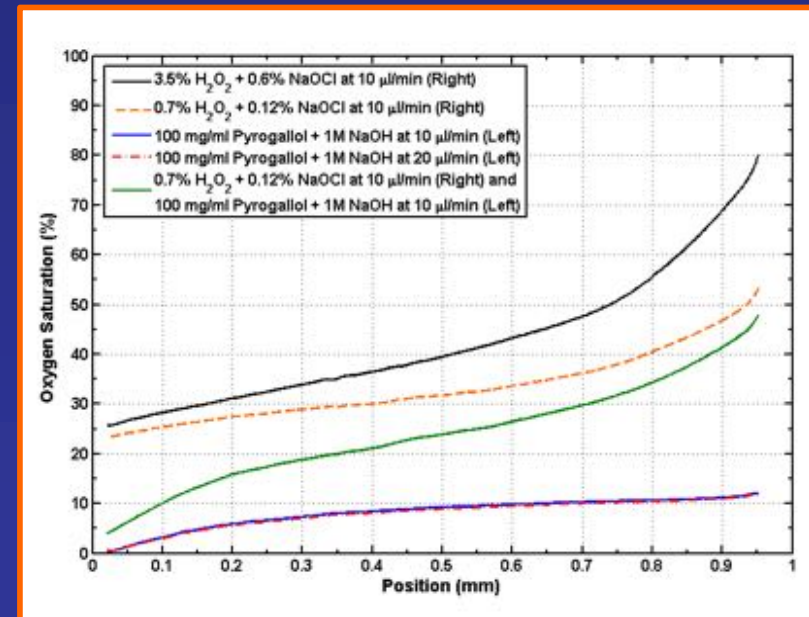
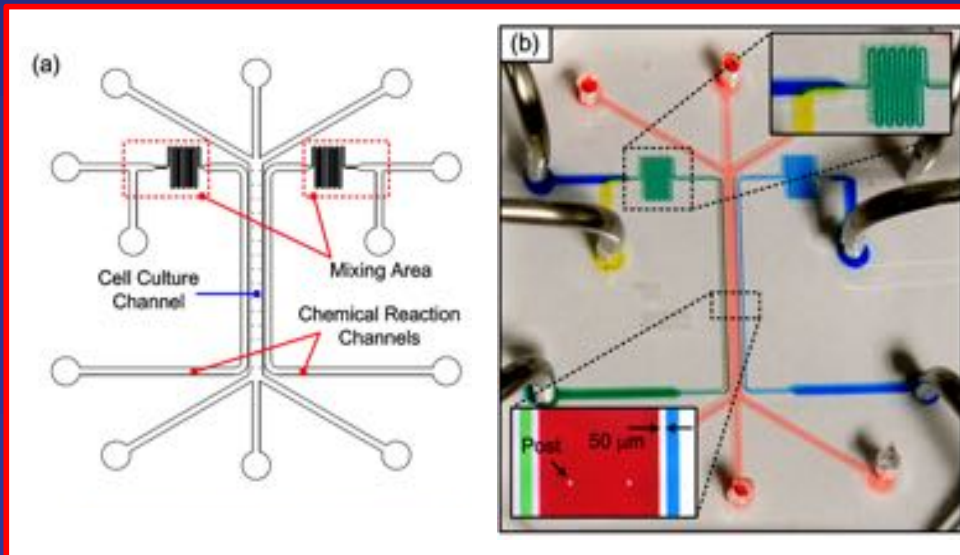
# Introduction – Conventional Methods

- Existing methods to control gaseous compositions for cell culture are often complicated, require gas cylinders, tedious interconnections, and sophisticated flow control systems.
- Direct chemical additions that may alter cell behaviors.
- WITHOUT Spatial Control.



# New Microfluidic Device Design

- Spatially-Confined Chemical Reaction Method
- The device is constructed using polydimethylsiloxane (PDMS).

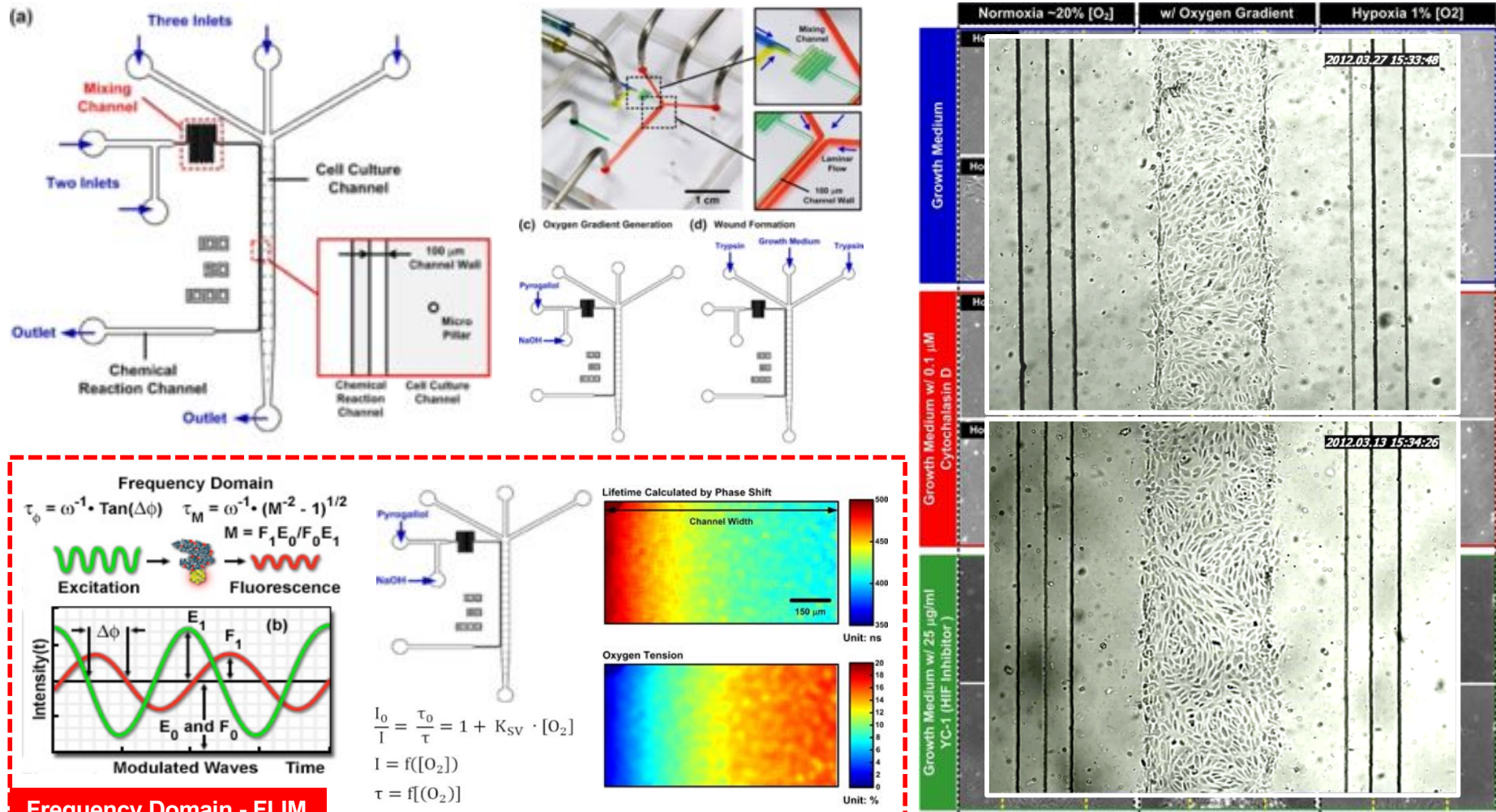


## Oxygen Diffusion Coefficient, D (cm<sup>2</sup>/s)

25°C in PDMS,  $D = 3.55 \times 10^{-5}$   
 20°C in Water,  $D = 1.97 \times 10^{-5}$   
 40°C in Water,  $D = 3.24 \times 10^{-5}$

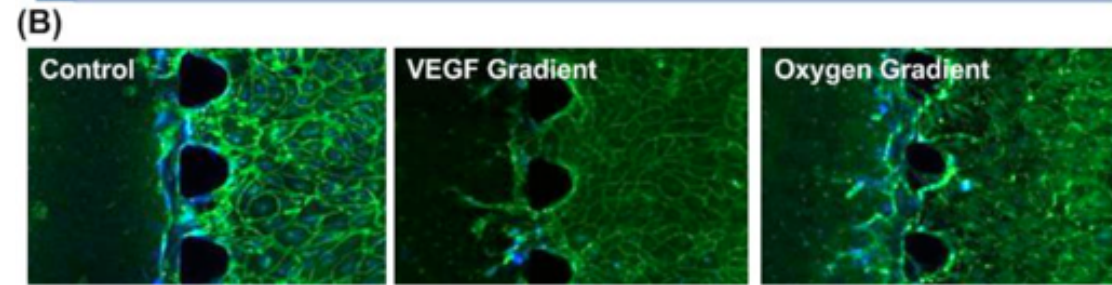
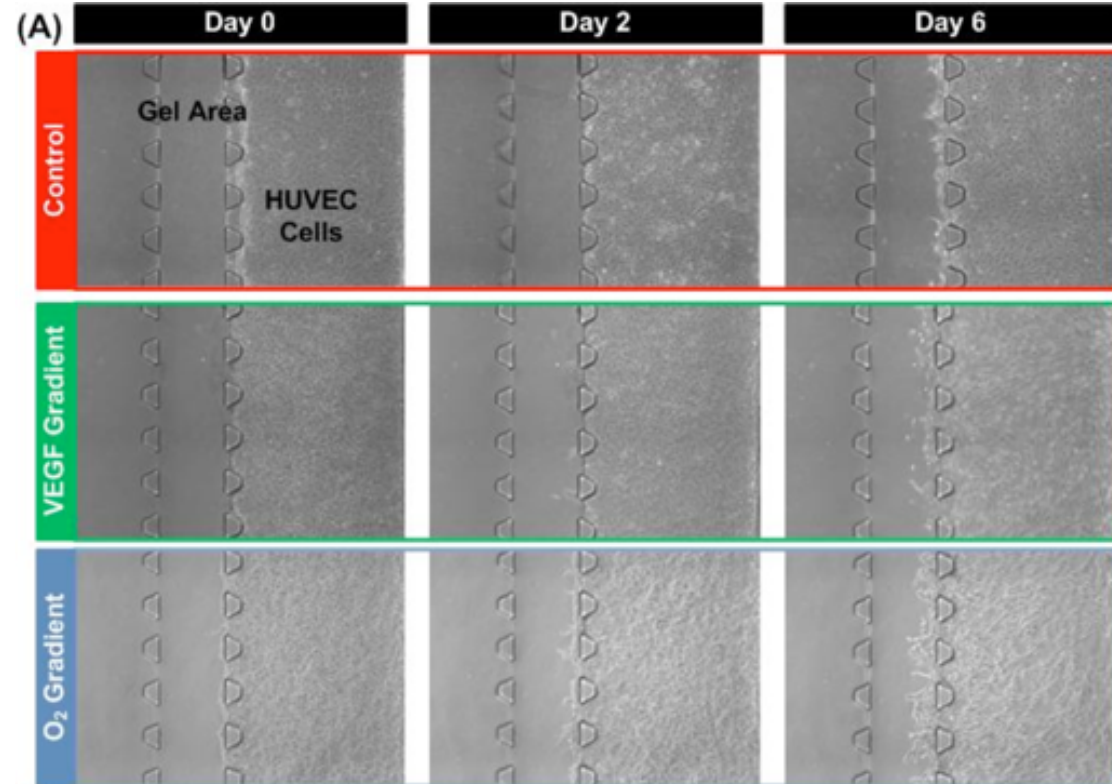
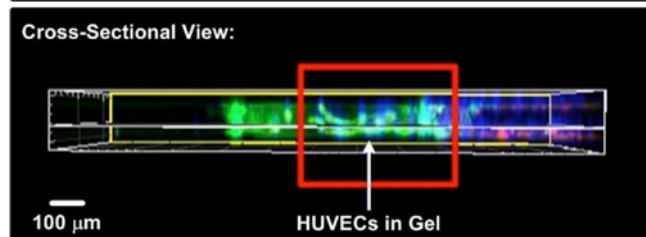
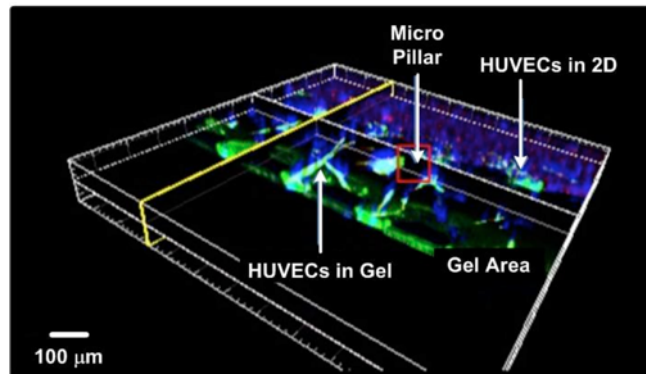
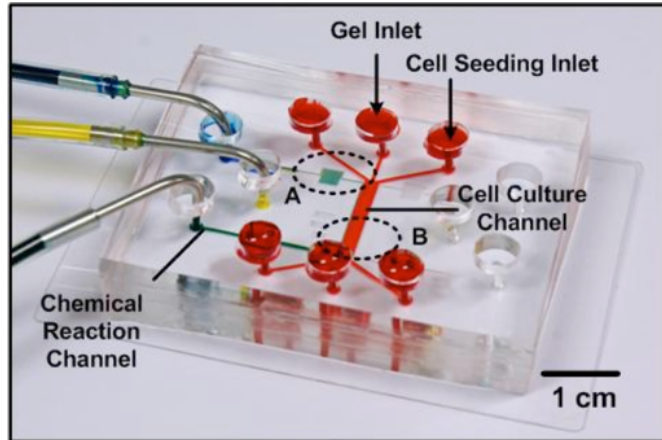
# Microfluidic Wound Healing Assay w/ O<sub>2</sub> Gradients

## Study Endothelial Cell Proliferation and Migration under Combinations of Oxygen Tensions, Gradients, and Drug Treatments



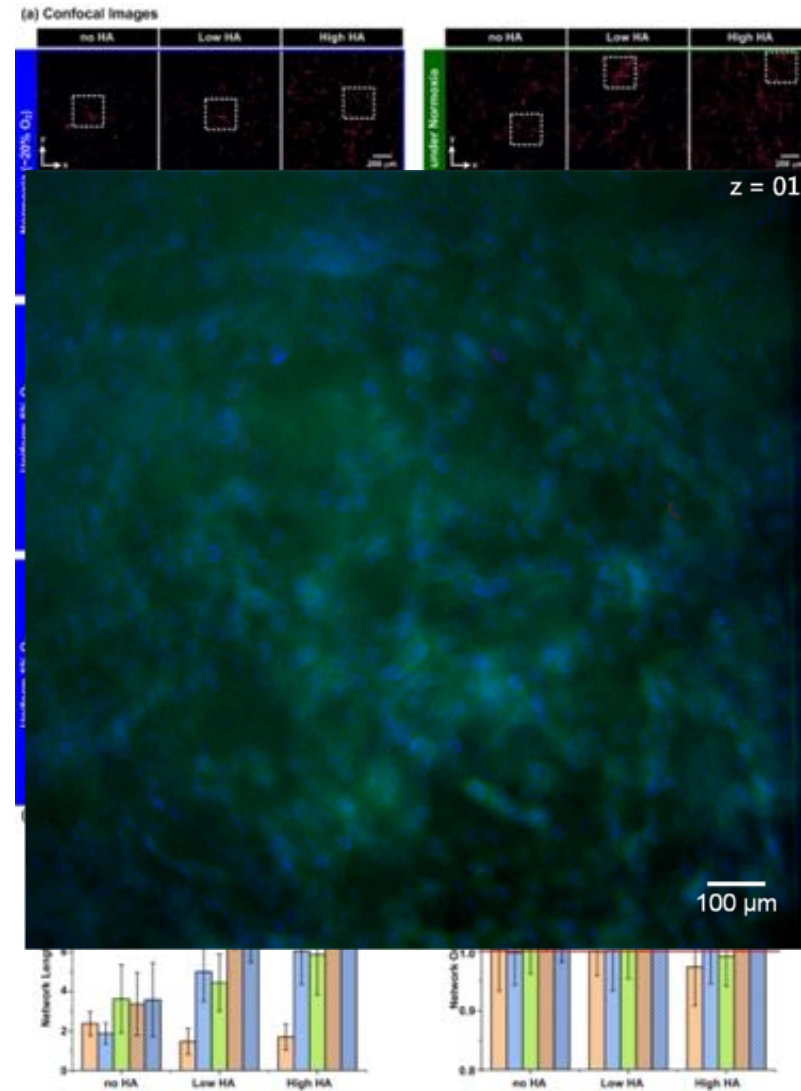
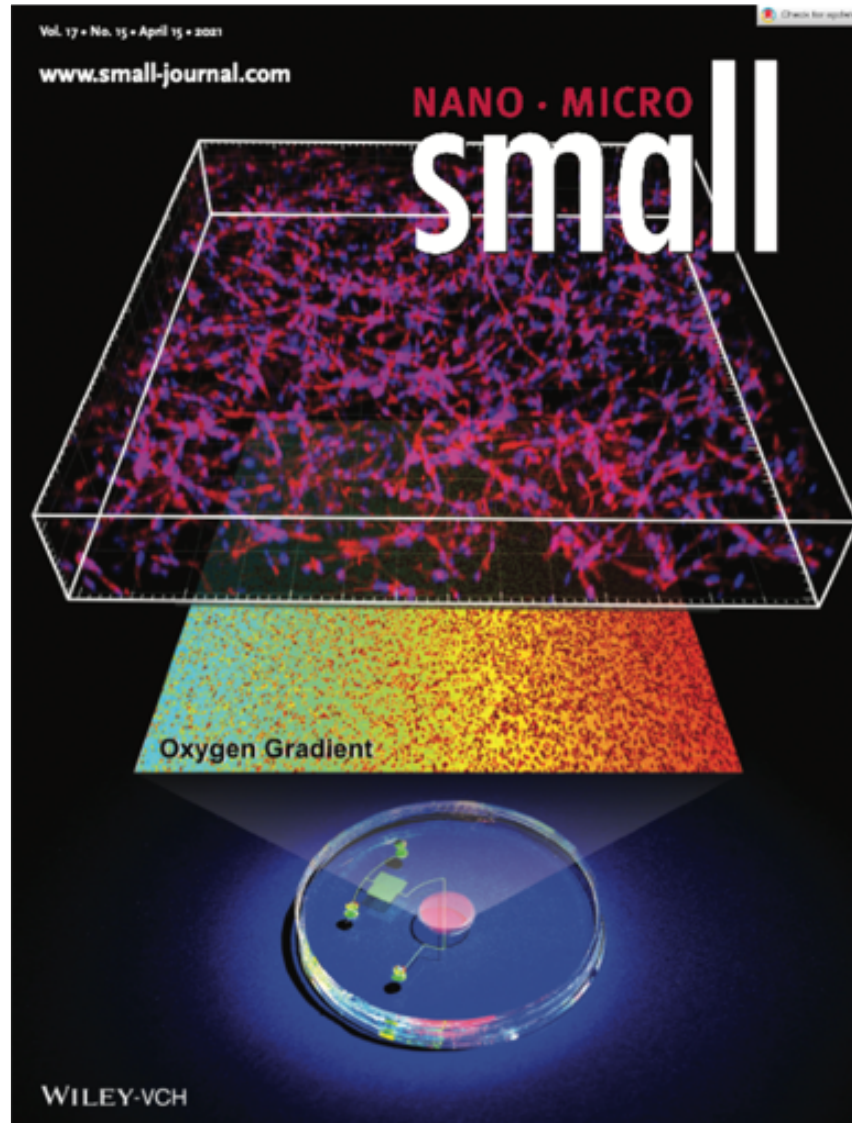
# Cell Sprouting in 3D under O<sub>2</sub> Gradients

## Study Endothelial Cell Sprouting in Three-Dimensional Matrix under Oxygen Gradients



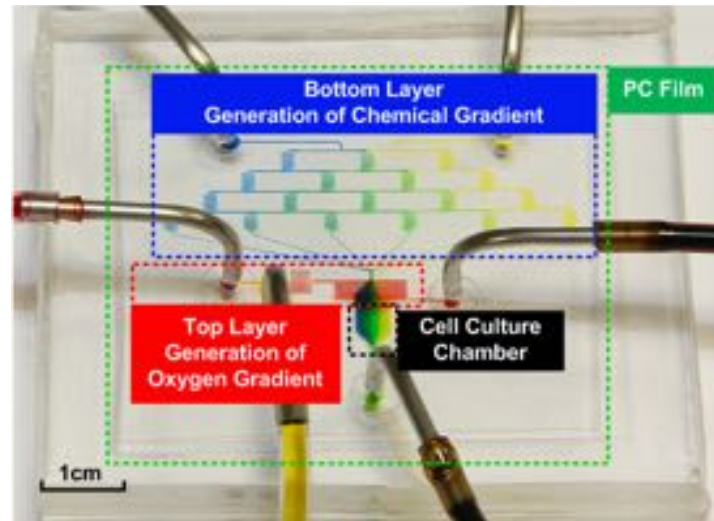
# Network Formation in 3D under O<sub>2</sub> Gradients

## Study 3D Network Formation of Endothelial Cells under Oxygen Gradients

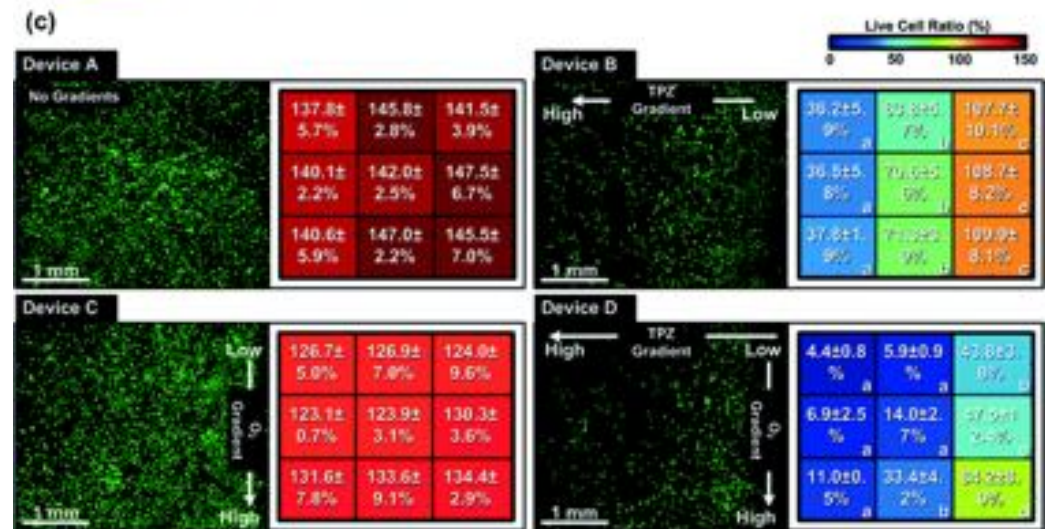
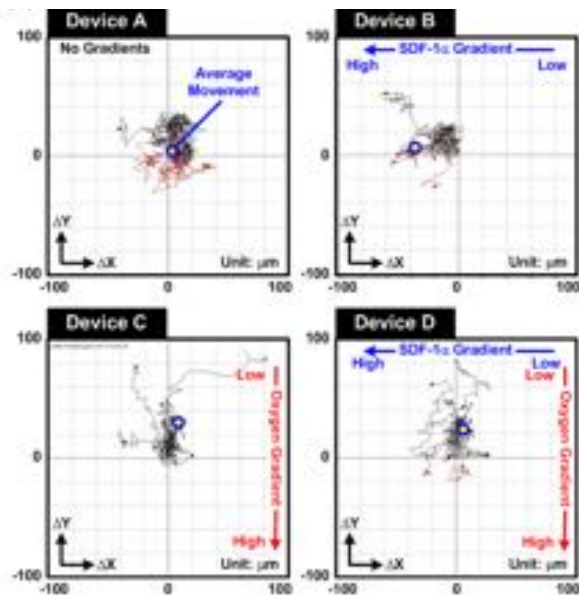
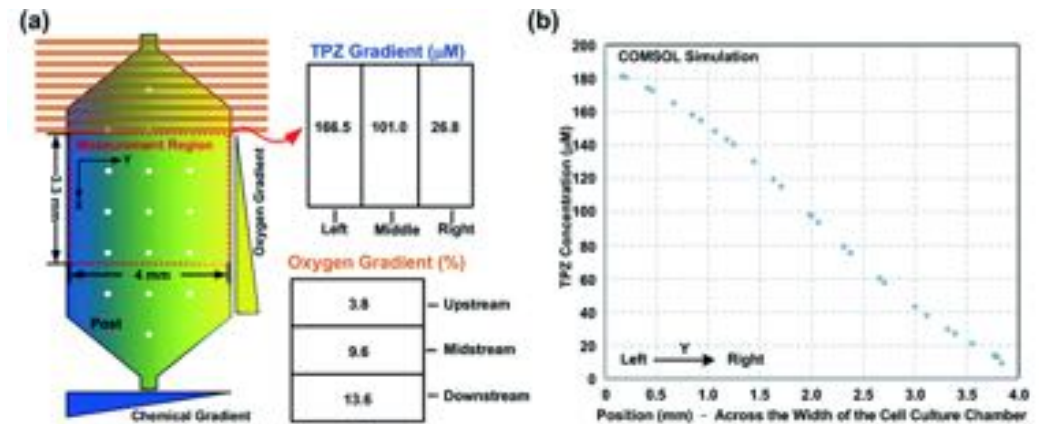


Accepted in *Small*.

# Microfluidic Devices for w/ O<sub>2</sub> and Chemical Gradients

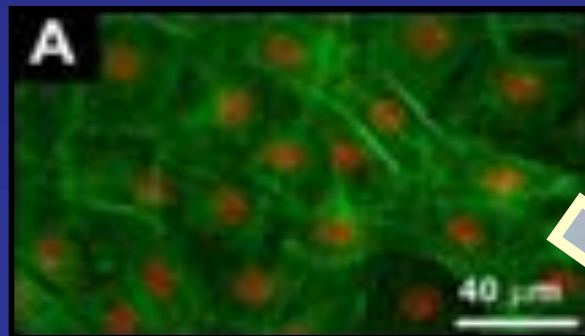


## Cell Culture Device w/ Chemical & O<sub>2</sub> Gradients (1 + 1 = 2 ?)



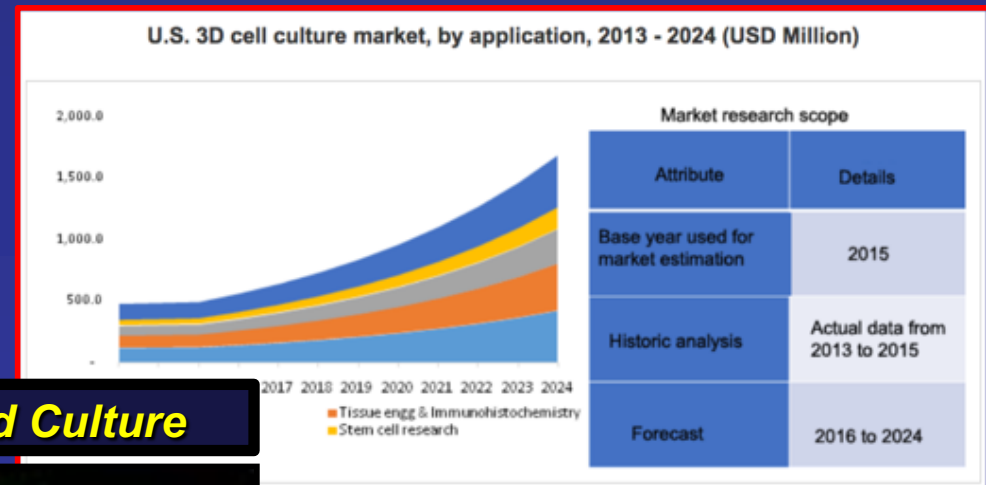
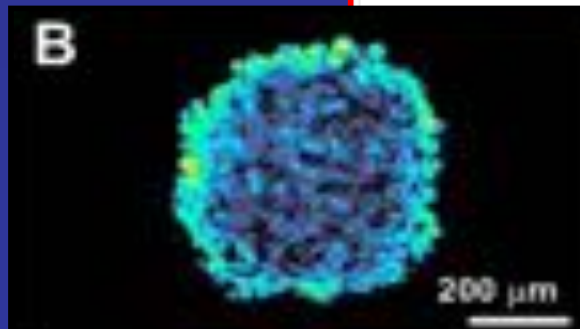
# Introduction – Additional Dimension

- Moving from cell monolayers to three-dimensional (3D) cultures is motivated by the need to work with cellular models that better mimic the environment of living tissues.
- For example, tumor spheroids have been widely used as an *in vitro* 3D model to simulate the multicellular microenvironment when investigating tumor cell physiology and responses to therapeutic agents.



**2D Monolayer**

**3D Spheroid Culture**

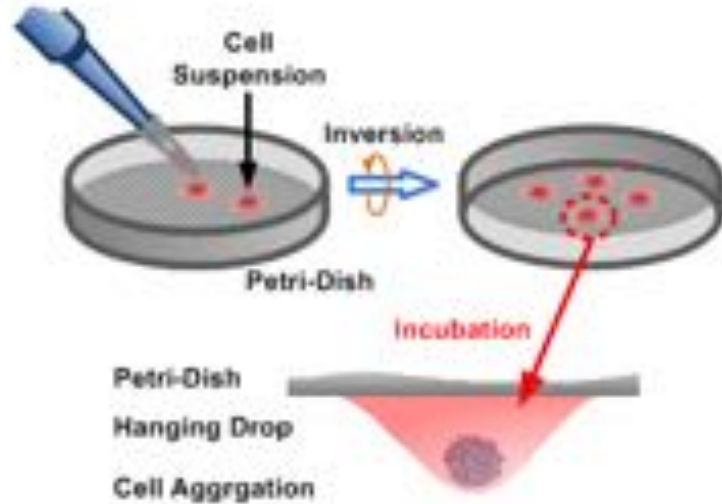


**Scaffold free 3D cell culture is expected to witness the fastest growth through to 2024.**



# Introduction - How to form Spheroids?

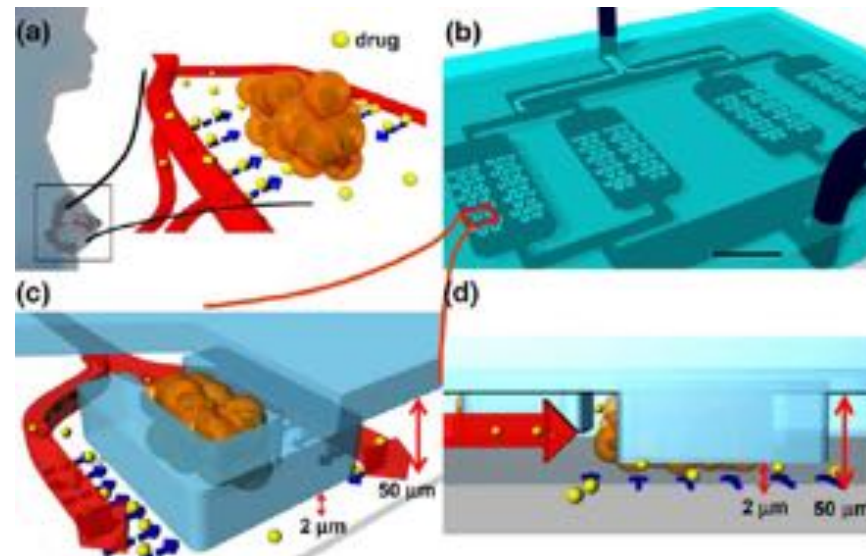
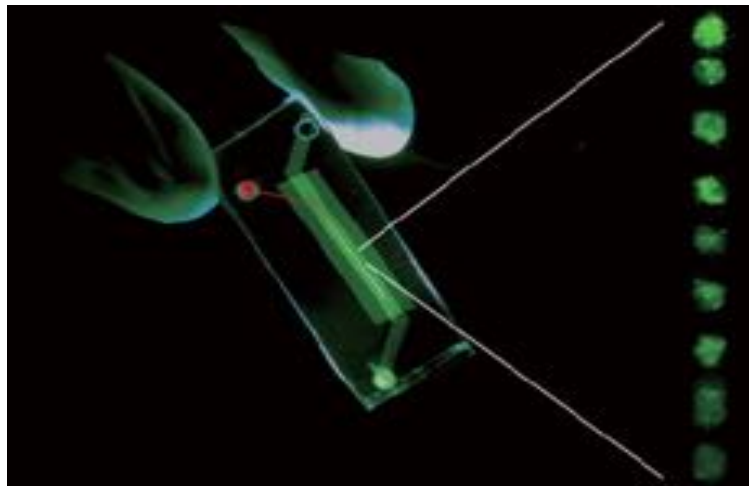
## Hanging Drop



## Rotating-Wall Vessel

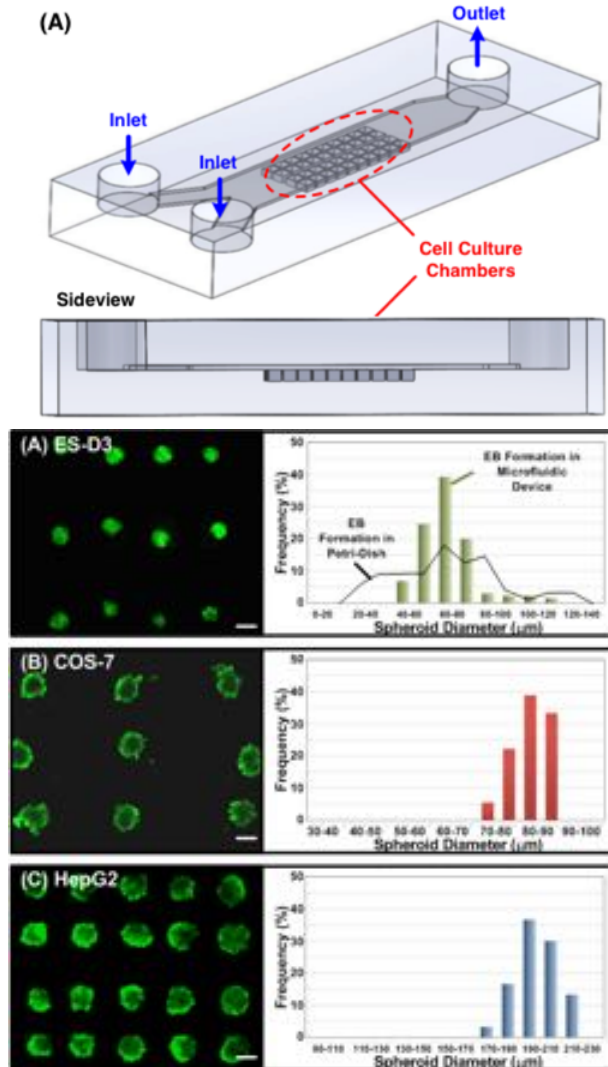


## Microfluidics



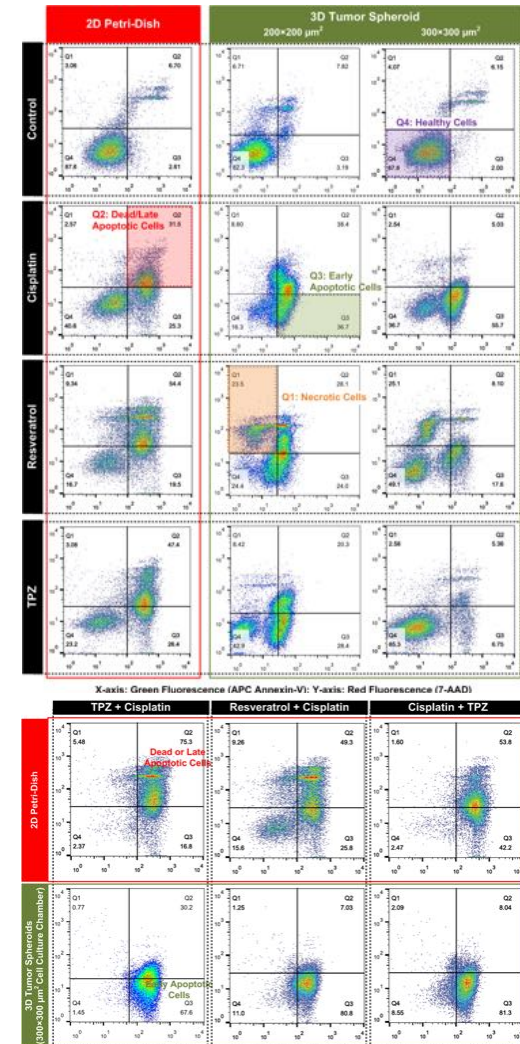
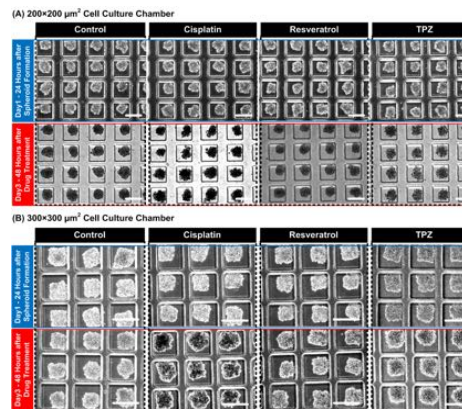
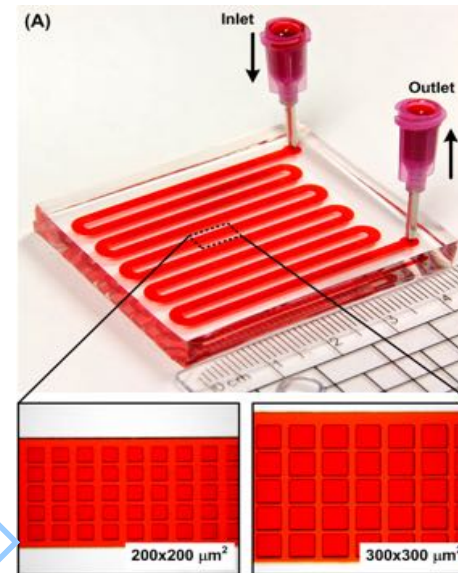
# Device for Spheroid Formation & Culture

## Uniform-Sized Spheroid Formation, Culture & Harvest



*Biomicrofluidics*, 2013, 7, 054114

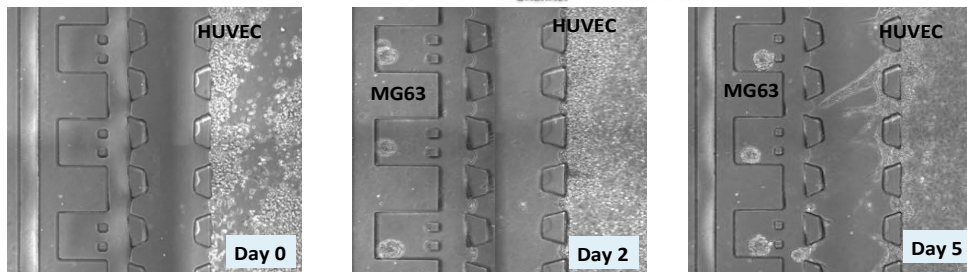
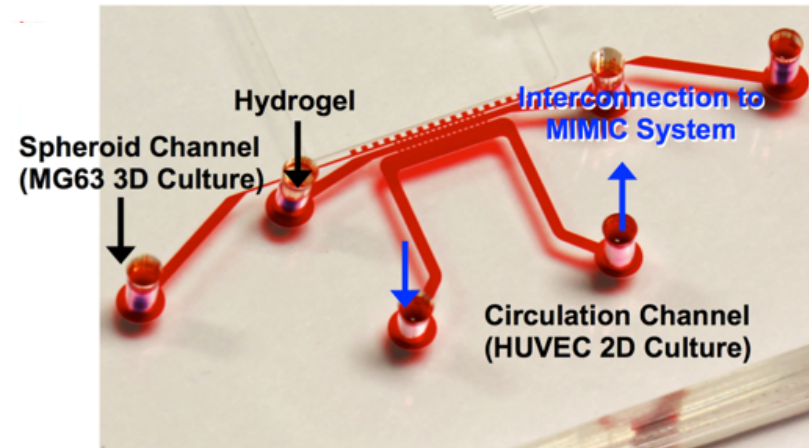
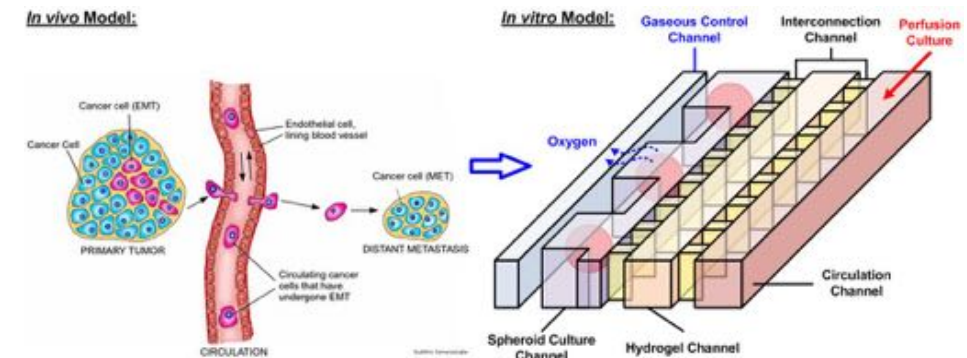
## Scale up (5000 Spheroids) for Flow Cytometry Analysis on Tumor Spheroids



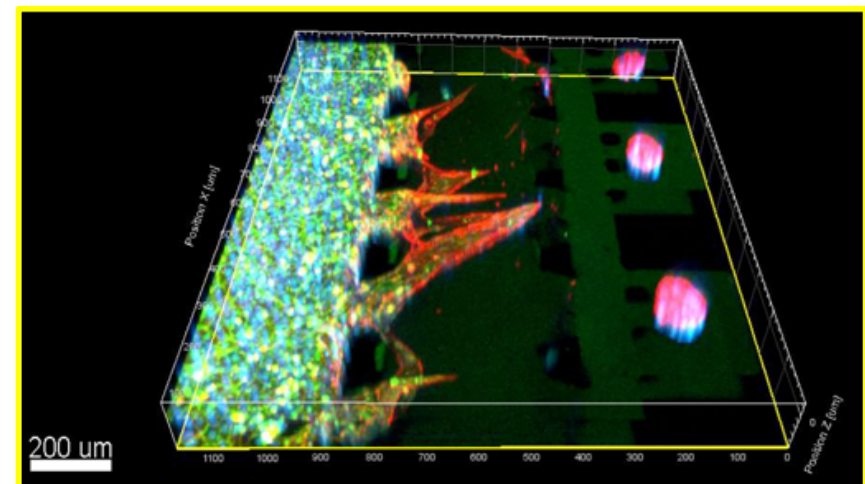
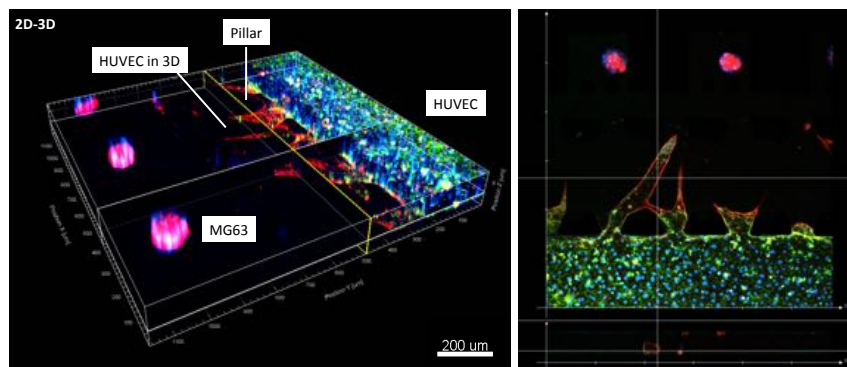
*Sci. Rep.*, 2016, 6, 21061

# Integrate Multiple Dimensions in a Platform

## 3D Osteosarcoma Cell (MG63) Spheroid and 2D Monolayer HUVEC Co-Culture to Study Endothelial Cell Sprouting into 3D Hydrogel Affected by Tumor Spheroids



Blue: DAPI Red: F-Actin Green: VE-Cadherin



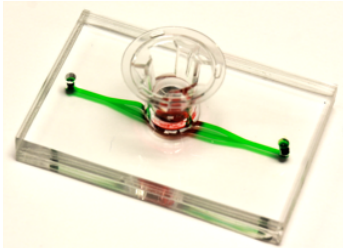
Presented in microTAS 2017, Manuscript in Preparation.

# Microfluidic *in vitro* Circulatory System

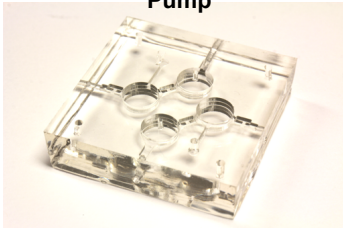
## Modularized Integrated Microfluidic (MIMIC) System to Mimic Circulatory Systems for *in vitro* Cell Studies

### MIMIC System – *in vitro* Circulatory System

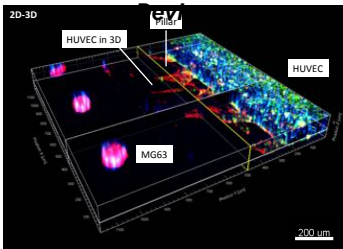
#### *In vitro* Blood-Air Barrier



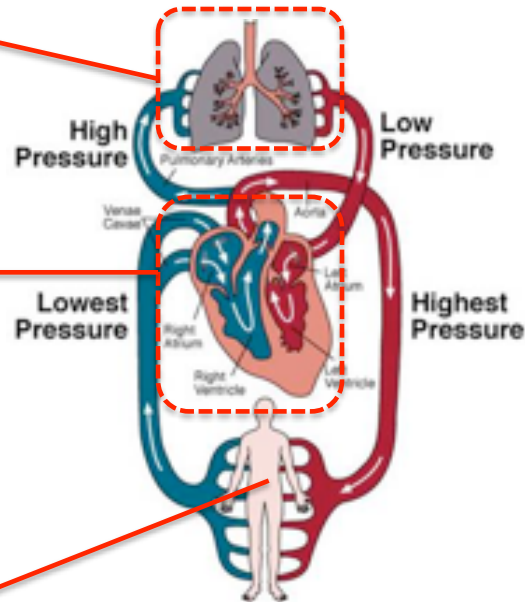
#### Heart-Mimicking Microfluidic Pump



#### Cell Co-Culture Microfluidic

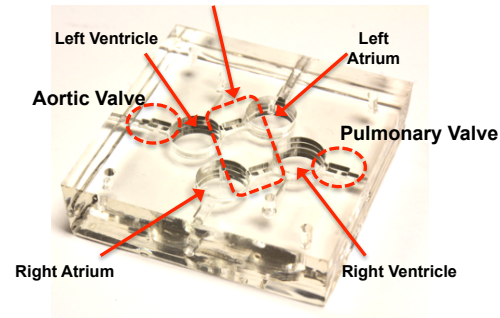


### *In vivo* Circulatory System

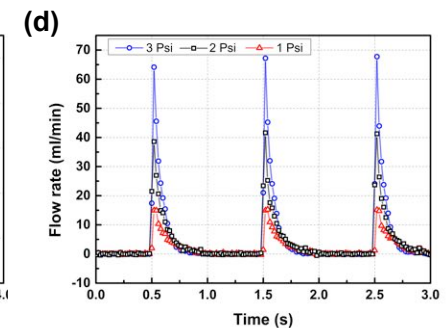
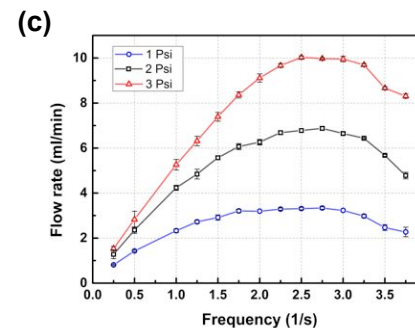
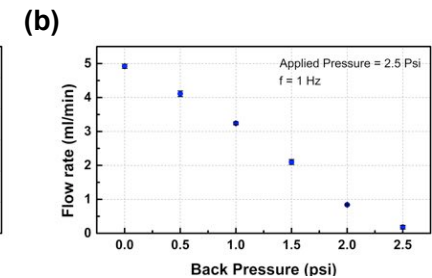
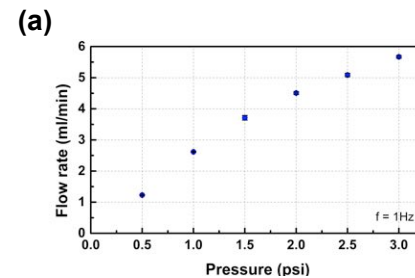
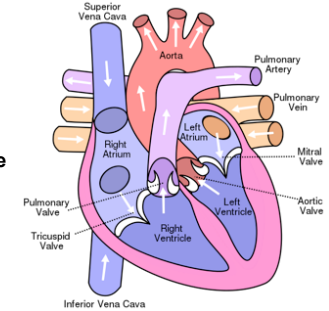


### Heart-Mimicking Microfluidic Pump

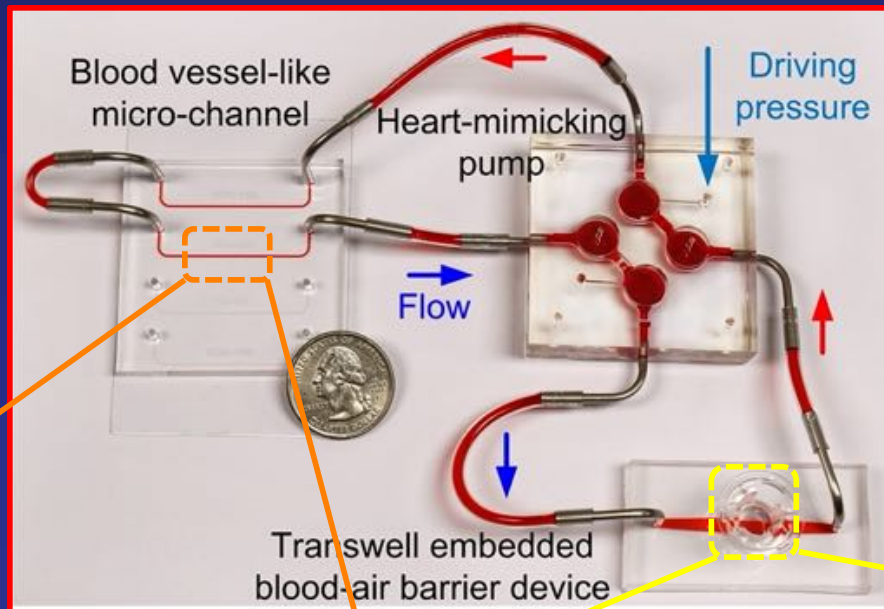
#### Atrioventricular Valves



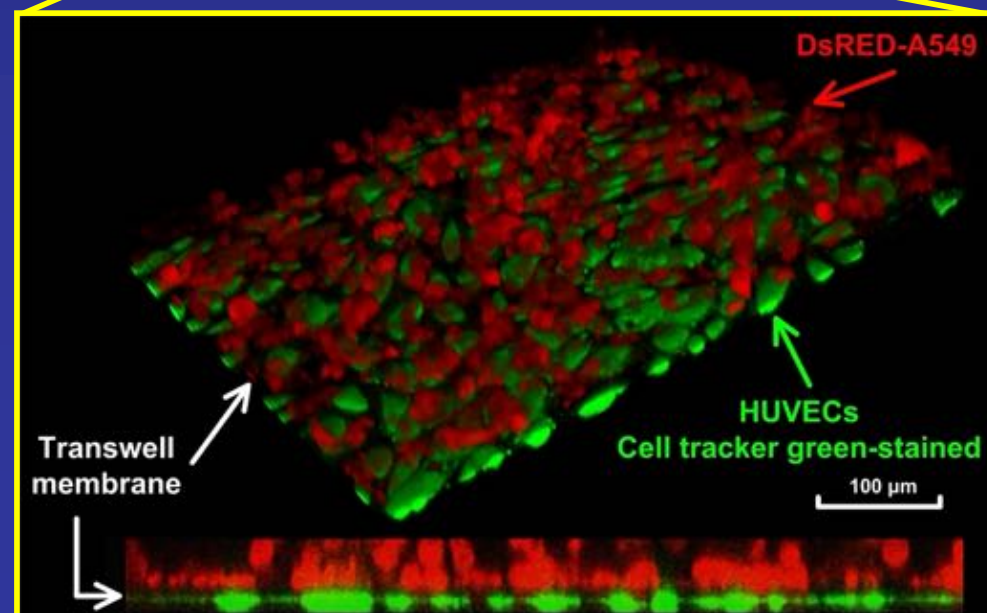
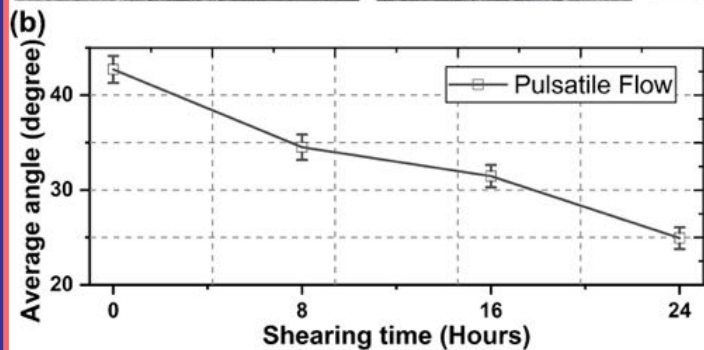
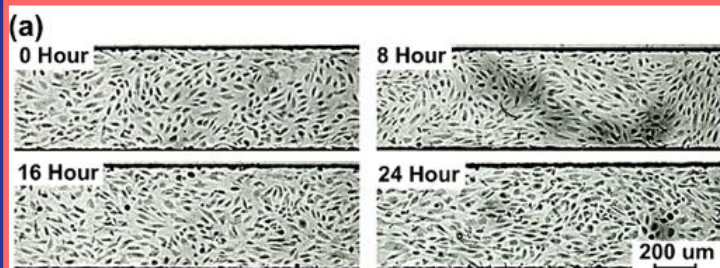
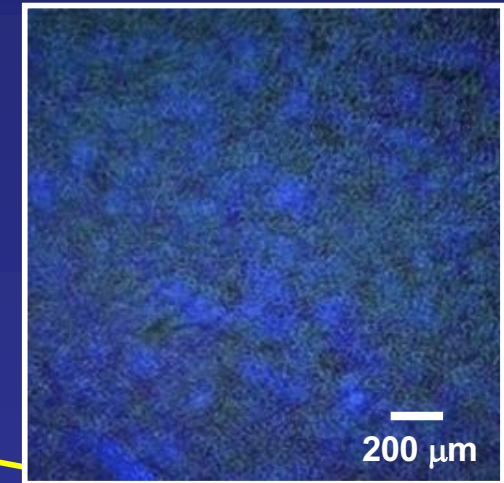
### Human Heart



# Toward Systemic Model – Integration



HUVEC/A549  
(Live/Dead Stain)



# Introduction – Engineering a Robot?

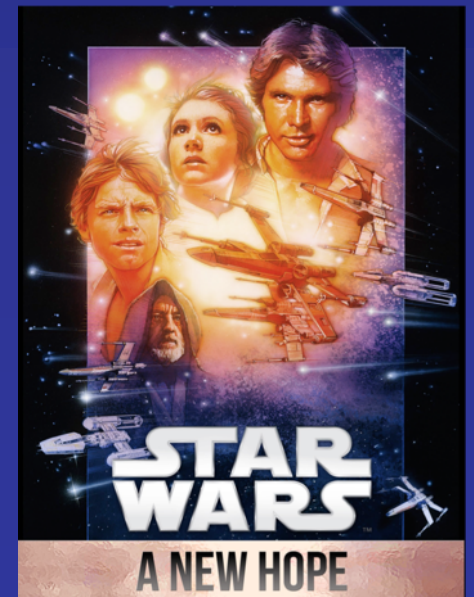
- Different Thought about Robot?



# Conclusion

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- Development of various microfluidic cell culture devices capable of controlling and analyzing gaseous and physical microenvironments.
- Multi-dimensional cell culture to mimic physiological arrangements of the cells *in vivo*.
- The new systemic *in vitro* model will provide biomedical scientists next generation cell culture models with better prediction power, and the results can be more easily to be translated into animal experiments and clinical tests.



# Acknowledgement

- Funding: MOST, NHRI, MOHW, BioTREC, Academia Sinica.
- Academic Collaboration (中研院、台大、陽明交通、北醫、國衛院、清華、慈濟大學、University of Michigan, Ohio State University, Canther.fr).
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- RCAS Colleagues.
- RCAS.TungLab Members.

